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Term: l12 and (PCR or polymerase chain reaction)

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L15	l12 and (PCR or polymerase chain reaction)	24	L15
L14	l12 and (PCR or polymease chain reaction)	18	L14
L13	L12 and cleav\$3 group	0	L13
L12	l10 and incorporat\$3	26	L12
L11	L10 and (cleav\$4 near5 incorporat\$3)	0	L11
L10	L9 and phosphate backbone	26	L10
L9	l4 and (chemic\$4 near5 cleav\$4)	88	L9
L8	L7 and (polymerase chain reactio or PCR)	23	L8
L7	l5 and (end\$1 or termin\$3)	44	L7
L6	L5 and PCR	23	L6
L5	L4 and (modif\$7 near5 sugar\$1)	44	L5
L4	L3 and (free near5 hydroxyl)	186	L4
L3	L2 and immobili\$7	2113	L3
L2	(primer\$1 or probe\$1 or oligonucleotide\$1) near5 cleav\$4	5099	L2
L1	6287821.pn.	2	L1

=> s (primer# or probe# or oligonucleotide#)(10a)immobiliz#####(10a)cleav####
 L1 85 (PRIMER# OR PROBE# OR OLIGONUCLEOTIDE#)(10A) IMMOBILIZ#####(10
 A) CLEAV####

=> s l1 and (free (10a)hydroxyl)
 L2 1 L1 AND (FREE (10A) HYDROXYL)

=> d l2 bib ab kwic

L2 ANSWER 1 OF 1 CAPLUS COPYRIGHT 2004 ACS on STN
 AN 1995:551055 CAPLUS
 DN 122:310286
 TI Tag reagent containing reporter groups identifiable by mass spectrometry
 and use of tag reagent in nucleic acid sequence determination
 IN Southern, Edwin; Cummins, William Jonathan
 PA ISIS Innovation Ltd., UK
 SO PCT Int. Appl., 67 pp.
 CODEN: PIXXD2
 DT Patent
 LA English
 FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9504160	A1	19950209	WO 1994-GB1675	19940801
	W: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, ES, FI, GB, GE, HU, JP, KE, KG, KP, KR, KZ, LK, LT, LU, LV, MD, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SI, SK, TJ, TT, UA, US, UZ, VN				
	RW: KE, MW, SD, AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
	CA 2168010	AA	19950209	CA 1994-2168010	19940801
	CA 2168010	C	20021001		
	AU 9472691	A1	19950228	AU 1994-72691	19940801
	AU 695349	B2	19980813		
	EP 711362	A1	19960515	EP 1994-922966	19940801
	EP 711362	B1	19971029		
	R: AT, BE, CH, DE, DK, ES, FR, GB, IT, LI, NL, SE				
	CN 1131440	A	19960918	CN 1994-193442	19940801
	CN 1088758	B	20020807		
	HU 73802	A2	19960930	HU 1996-27	19940801
	JP 09501830	T2	19970225	JP 1994-505687	19940801
	EP 778280	A2	19970611	EP 1996-119962	19940801
	EP 778280	A3	19990127		
	EP 778280	B1	20030102		
	R: AT, BE, CH, DE, DK, ES, FR, GB, IT, LI, NL, SE				
	AT 159767	E	19971115	AT 1994-922966	19940801
	ES 2108479	T3	19971216	ES 1994-922966	19940801
	RU 2158310	C2	20001027	RU 1996-103646	19940801
	JP 3289911	B2	20020610	JP 1995-505687	19940801
	AT 230409	E	20030115	AT 1996-119962	19940801
	FI 9600403	A	19960129	FI 1996-403	19960129
	NO 9600370	A	19960328	NO 1996-370	19960129
	US 5770367	A	19980623	US 1996-586875	19960205
	US 6218111	B1	20010417	US 1997-988384	19971210
	US 2001031472	A1	20011018	US 2001-810162	20010319
	US 2002115091	A1	20020822	US 2001-3830	20011206
	US 6576426	B2	20030610		
PRAI	GB 1993-15847	A	19930730		
	EP 1994-922966	A3	19940801		
	WO 1994-GB1675	W	19940801		
	US 1996-586875	A1	19960205		
	US 1997-988384	A3	19971210		
	US 2001-810162	B1	20010319		
AB	A reagent A-L-R (A=an analyte moiety comprising at least two analyte residues; L=a linking moiety; R=a tag moiety comprising one or more				

reporter groups adapted for detection by mass spectrometry) is described. The reporter group at each position of the tag moiety is chosen to designate an analyte residue at a defined position of the analyte moiety. A plurality of such reagents, each comprising a different analyte moiety, provides a library of reagents which may be used in assay methods involving a target substance. Anal. of the tag moieties indicates the nature of the analyte moieties bound to the target substance. A method of sequencing nucleic acid employs a library of the reagents to determine the sequence of a target nucleic acid. One A-L-R reagent precursor (I) was prepared. This precursor contains a dimethoxytrityl group for attachment of the oligonucleotide analyte, an O-t-butyldiphenylsilyl group for attachment of a tag, an N-hydroxysuccinimide group for attachment of the precursor to a solid support (to facilitate attachment of tag and analyte), and a tertiary amine group for conversion to a pos. charged group to mass spectrometric anal. The o-nitrophenol group provides for photocleavage of the A-L-R reagent. I was used to synthesize a tagged oligonucleotide which was employed in nucleic acid sequence determination by progressive ligation. The sequencing method comprised: (1) hybridization of the target sequence to an immobilized oligonucleotide; (2) incubation of the immobilized target with a library of tagged oligonucleotides; (3) ligase-mediated attachment of the hybridized tagged oligonucleotide with the immobilized oligonucleotide; (4) removal of nonligated tagged oligonucleotides; and (5) photolytic cleavage of the linker to release the tag (which is analyzed by mass spectrometry) and to generate a free 3' hydroxyl which can be further probed with tagged oligonucleotides.

AB A reagent A-L-R (A=an analyte moiety comprising at least two analyte residues; L=a linking moiety; R=a tag moiety comprising one or more reporter groups adapted for detection by mass spectrometry) is described. The reporter group at each position of the tag moiety is chosen to designate an analyte residue at a defined position of the analyte moiety. A plurality of such reagents, each comprising a different analyte moiety, provides a library of reagents which may be used in assay methods involving a target substance. Anal. of the tag moieties indicates the nature of the analyte moieties bound to the target substance. A method of sequencing nucleic acid employs a library of the reagents to determine the sequence of a target nucleic acid. One A-L-R reagent precursor (I) was prepared. This precursor contains a dimethoxytrityl group for attachment of the oligonucleotide analyte, an O-t-butyldiphenylsilyl group for attachment of a tag, an N-hydroxysuccinimide group for attachment of the precursor to a solid support (to facilitate attachment of tag and analyte), and a tertiary amine group for conversion to a pos. charged group to mass spectrometric anal. The o-nitrophenol group provides for photocleavage of the A-L-R reagent. I was used to synthesize a tagged oligonucleotide which was employed in nucleic acid sequence determination by progressive ligation. The sequencing method comprised: (1) hybridization of the target sequence to an immobilized oligonucleotide; (2) incubation of the immobilized target with a library of tagged oligonucleotides; (3) ligase-mediated attachment of the hybridized tagged oligonucleotide with the immobilized oligonucleotide; (4) removal of nonligated tagged oligonucleotides; and (5) photolytic cleavage of the linker to release the tag (which is analyzed by mass spectrometry) and to generate a free 3' hydroxyl which can be further probed with tagged oligonucleotides.

=> s l1 and (chemic####(10a)cleav####)
L3 3 L1 AND (CHEMIC####(10A) CLEAV####)

=> d l3 1-3 bib ab kwic

L3 ANSWER 1 OF 3 CAPLUS COPYRIGHT 2004 ACS on STN

AN 2003:750104 CAPLUS
 DN 139:208379
 TI **Chemical cleavage** reactions of DNA on solid support:
 application in mutation detection
 AU Bui, Chinh T.; Lambrinakos, Andreana; Babon, Jeffrey J.; Cotton, Richard
 G. H.
 CS Genomic Disorders Research Centre, St. Vincent's Hospital, University of
 Melbourne, Fitzroy, 3065, Australia
 SO BMC Chemical Biology (2003), 3, No pp. given
 CODEN: BCBMBZ; ISSN: 1472-6769
 URL: <http://www.biomedcentral.com/1472-6769/3/1>
 PB BioMed Central Ltd.
 DT Journal; (online computer file)
 LA English
 AB Background: The conventional solution-phase Chemical Cleavage of Mismatch (CCM)
 method is time-consuming, as the protocol requires purification of DNA after
 each reaction step. This paper describes a new version of CCM to overcome
 this problem by immobilizing DNA on silica solid supports. Results: DNA
 test samples were loaded on to silica beads and the DNA bound to the solid
 supports underwent chemical modification reactions with KMnO4 (potassium
 permanganate) and hydroxylamine in 3M TEAC (tetraethylammonium chloride)
 solution. The resulting modified DNA was then simultaneously cleaved by
 piperidine and removed from the solid supports to afford DNA fragments
 without the requirement of DNA purification between reaction steps.
 Conclusions: The new solid-phase version of CCM is a fast, cost-effective
 and sensitive method for detection of mismatches and mutations.

RE.CNT 13 THERE ARE 13 CITED REFERENCES AVAILABLE FOR THIS RECORD
 ALL CITATIONS AVAILABLE IN THE RE FORMAT

TI **Chemical cleavage** reactions of DNA on solid support:
 application in mutation detection
 IT **Probes** (nucleic acid)
 RL: ARG (Analytical reagent use); ANST (Analytical study); USES (Uses)
 (immobilized on silica beads; chemical **cleavage**
 reactions of DNA on solid support and application to mismatch mutation
 detection)
 IT 7631-86-9, Silica, analysis
 RL: ARU (Analytical role, unclassified); DEV (Device component use); ANST
 (Analytical study); USES (Uses)
 (DNA **probe-immobilized** beads; chemical
cleavage reactions of DNA on solid support and application to
 mismatch mutation detection)

L3 ANSWER 2 OF 3 CAPLUS COPYRIGHT 2004 ACS on STN

AN 1998:728141 CAPLUS

DN 129:326948

TI Sizing of primer extension products using primers containing a
chemically cleavable linkage

IN Monforte, Joseph Albert; Becker, Christopher Hank; Shaler, Thomas Andrew;
 Pollart, Daniel Joseph

PA SRI International, USA

SO U.S., 59 pp., Cont.-in-part of U.S. Ser. No. 445,751.

CODEN: USXXAM

DT Patent

LA English

FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 5830655	A	19981103	US 1996-639363	19960426
	US 5700642	A	19971223	US 1995-445751	19950522
	CA 2220418	AA	19961128	CA 1996-2220418	19960430
	CA 2220418	C	20030603		
	WO 9637630	A1	19961128	WO 1996-US6116	19960430
	W: AU, CA, CN, JP, KR				
	RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				

AU 9656352	A1	19961211	AU 1996-56352	19960430
AU 695705	B2	19980820		
EP 828855	A1	19980318	EP 1996-913305	19960430
EP 828855	B1	19991215		
R: DE, FR, GB, IT, NL				
CN 1191575	A	19980826	CN 1996-195707	19960430
JP 11505127	T2	19990518	JP 1996-535680	19960430
JP 3437184	B2	20030818		
PRAI US 1995-445751	A2	19950522		
US 1996-639363	A	19960426		
WO 1996-US6116	W	19960430		

AB A method for sizing oligonucleotide primer extension products that minimizes the effect of the primer size on the determination is described. The method can be used to detect mutations, e.g. in mol. diagnosis. The method uses immobilized primers that have an unusual linkage in the phosphodiester backbone near the 3'-end of the primer that can be cleaved chemical Upon selective cleavage of the cleavable site, primer extension products that contain ≤ 5 bases from the primer are released, to provide more useful sizing and sequence information per fragment than extension products containing the entire primer. Methods for the synthesis of oligonucleotides carrying a modified internucleoside linkage such as a dialkoxysilane or a phosphorothioate is described. Primers containing a riboside that creates a base-cleavable linkage are also described.

RE.CNT 44 THERE ARE 44 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

TI Sizing of primer extension products using primers containing a **chemically cleavable** linkage

IT **Immobilization**, biochemical
(of **primers** containing **cleavable** linkages; sizing of **primer** extension products using primers containing chemical **cleavable** linkage)

L3 ANSWER 3 OF 3 BIOSIS COPYRIGHT (c) 2004 The Thomson Corporation. on STN
AN 2000:369544 BIOSIS
DN PREV200000369544
TI Method for the quantitative detection of specific nucleic acid sequences.
AU Leying, Hermann [Inventor, Reprint author]; Hinzpeter, Matthias [Inventor]; Wittor, Heiko [Inventor]; Fritton, Hans-Peter [Inventor]
CS Bichl, Germany
ASSIGNEE: Roche Diagnostics GmbH, Mannheim, Germany
PI US 6027886 February 22, 2000
SO Official Gazette of the United States Patent and Trademark Office Patents, (Feb. 22, 2000) Vol. 1231, No. 4. e-file.
CODEN: OGUPE7. ISSN: 0098-1133.
DT Patent
LA English
ED Entered STN: 30 Aug 2000
Last Updated on STN: 8 Jan 2002

AB Method and kit for the quantitative detection of specific oligonucleotide or polynucleotide sequences which is characterized in that a sample mixture containing RNA or single-stranded DNA is hybridized with an oligonucleotide or polynucleotide probe(s) which are complementary to the nucleotide sequence to be determined and carry(ies) a specifically bindable and a detectable **chemical** group subsequently it is admixed with an agent that **cleaves** single-stranded polynucleotide sequences and the **immobilized** or non-**immobilized** nucleotide **probe** is determined after transfer into a suitable reaction vessel. It has proven to be particularly advantageous when a mixture is used which is composed of different cleaving reagents.

AB. . . polynucleotide probe(s) which are complementary to the nucleotide sequence to be determined and carry(ies) a specifically bindable and a detectable **chemical** group subsequently it is admixed with an agent that **cleaves** single-stranded polynucleotide sequences and

the **immobilized** or non-**immobilized** nucleotide
probe is determined after transfer into a suitable reaction
vessel. It has proven to be particularly advantageous when a mixture is.

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